



Open Access

Diode Laser—Can It Replace the Electrical Current Used in Endoscopic Submucosal Dissection?

Yunho Jung¹, Gwang Ho Baik², Weon Jin Ko³, Bong Min Ko⁴, Seong Hwan Kim⁵, Jin Seok Jang⁶, Jae-Young Jang⁷, Wan-Sik Lee⁸, Young Kwan Cho⁵, Sun Gyo Lim⁹, Hee Seok Moon¹⁰, In Kyung Yoo¹¹ and Joo Young Cho¹¹

¹Department of Internal Medicine, Soonchunhyang University College of Medicine, Cheonan, ²Department of Internal Medicine, Hallym University College of Medicine, Chuncheon, ³Department of Internal Medicine, Inha University School of Medicine, Incheon, ⁴Department of Internal Medicine, Soonchunhyang University College of Medicine, Bucheon, ⁵Department of Internal Medicine, Nowon Eulji Medical Center, Eulji University College of Medicine, Seoul, ⁶Department of Internal Medicine, Dong-A University College of Medicine, Busan, ⁷Department of Gastroenterology and Hepatology, College of Medicine, Kyung Hee University, Seoul, ⁸Department of Internal Medicine, Chonnam National University Hwasun Hospital, Gwangju, ⁹Department of Gastroenterology, Ajou University School of Medicine, Suwon, ¹⁰Department of Internal Medicine, Chungnam National University Hospital, Daejeon, ¹¹Division of Gastroenterology, Department of Internal Medicine, CHA Bundang Medical Center, Seongnam, Korea

Background/Aims: A new medical fiber-guided diode laser system (FDLS) is expected to offer high-precision cutting with simultaneous hemostasis. Thus, this study aimed to evaluate the feasibility of using the 1,940-nm FDLS to perform endoscopic submucosal dissection (ESD) in the gastrointestinal tract of an animal model.

Methods: In this prospective animal pilot study, gastric and colorectal ESD using the FDLS was performed in *ex vivo* and *in vivo* porcine models. The completeness of *en bloc* resection, the procedure time, intraoperative bleeding, histological injuries to the muscularis propria (MP) layer, and perforation were assessed.

Results: The *en bloc* resection and perforation rates in the *ex vivo* study were 100% (10/10) and 10% (1/10), respectively; those in the *in vivo* study were 100% (4/4) and 0% for gastric ESD and 100% (4/4) and 25% (1/4) for rectal ESD, respectively. Deep MP layer injuries tended to occur more frequently in the rectal than in the gastric ESD cases, and no intraoperative bleeding occurred in either group.

Conclusions: The 1,940-nm FDLS was capable of yielding high *en bloc* resection rates without intraoperative bleeding during gastric and colorectal ESD in animal models. **Clin Endosc 2021;54:555-562**

Key Words: Endoscopic mucosal resection; Hemostasis; Lasers; Perforation

INTRODUCTION

Endoscopic submucosal dissection (ESD) is an advanced endoscopic technique used to remove superficial gastrointestinal (GI) tract neoplasms. This technique has advantages

enabling *en bloc* resection regardless of the tumor size.¹ The resection margins of tissue removed *en bloc* can be accurately evaluated in all directions by pathologists. Thus, local recurrence can be reduced by determining whether the lesions are completely resected.² Despite these advantages, ESD is not popular worldwide because of technical difficulties and high-risk complications. Thus, various types of endoscopic knives have been developed to overcome these technical difficulties; these include insulated tip knife-type (IT knife, IT knife 2, and IT knife nano), needle knife-type (hook knife, dual knife, flex knife, B-knife, and hybrid knife), and grasping scissor knife-type (SB knife, SB knife Jr, and Clutch cutter knife).^{1,3} These endoscopic knives have helped remove lesions more safely and effectively but can cause intraoperative or postoperative bleeding because of the electrosurgical current.

Received: August 19, 2020 **Revised:** October 9, 2020

Accepted: October 15, 2020

Correspondence: Joo Young Cho

Division of Gastroenterology, Department of Internal Medicine, CHA Bundang Medical Center, 59 Yatap-ro, Bundang-gu, Seongnam 13496, Korea

Tel: +82-31-780-5641, **Fax:** +82-32-780-5005, **E-mail:** cjoy6695@naver.com

ORCID: <https://orcid.org/0000-0002-7182-5806>

© This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

Bleeding is one of the most common complications associated with ESD, as intraprocedural bleeding interferes with precise endoscopic resection under a clear view of the operating field, which can lead to longer procedure times or an increase in the frequency of complications, such as perforation.⁴ The incidence rates of immediate and delayed bleeding associated with gastric ESD are approximately 22% and 4.5%–5.5%, respectively.^{5–7} A recent meta-analysis showed that the incidence rates of immediate and delayed bleeding associated with colorectal ESD are 0.82% and 1.7%, respectively.⁸

Lasers generate heat and cause edema in the vessel wall, resulting in hemostasis.⁹ Therefore, attempts have been made to reduce the risk of bleeding associated with these laser properties by using several types of lasers, including carbon dioxide (CO₂) laser,^{10,11} Nd: YAG laser,¹² thulium laser,¹³ and diode laser,¹⁴ for endoscopic procedures.

The diode laser system has been used as a surgical instrument for incising and dissecting tissue and for achieving hemostasis. However, no study has yet performed ESD using the 1,940-nm diode laser system. A new medical fiber-guided diode laser system (FDLS) designed for endoscopic procedures emitting a high-frequency 1,940-nm wavelength is expected to offer high-precision cutting with simultaneous hemostasis. Thus, this study aimed to evaluate the feasibility of the application of the FDLS for performing ESD in the GI tract of an animal model.

MATERIALS AND METHODS

This study was designed as a prospective animal pilot study. Herein, the feasibility of endoscopic application of the FDLS in ESD was evaluated in *ex vivo* and *in vivo* porcine models. An *ex vivo* porcine model was used first to confirm the feasibility of FDLS application for ESD, followed by an *in vivo* porcine model to assess its clinical applicability.

Instruments

A single-channel endoscope (GIF-Q260J; Olympus Medical Systems Co., Tokyo, Japan) was used for all ESD procedures. A transparent distal cap (Olympus EndoTherapy; Olympus America Inc., Waltham, MA, USA) was attached to the tip of the endoscope to optimize visualization. A hypertonic saline solution mixed with diluted epinephrine and traces of indigo carmine were injected to lift the mucosal and submucosal layers.

A laser system (GI Laser; Wontech, Daejeon, Korea) emitting a high-frequency diode laser (1,000 Hz) and 1,940-nm wavelength was used for ESD. A specially designed fiber optic laser catheter (GI Laser Knife; Wontech) was used to protect

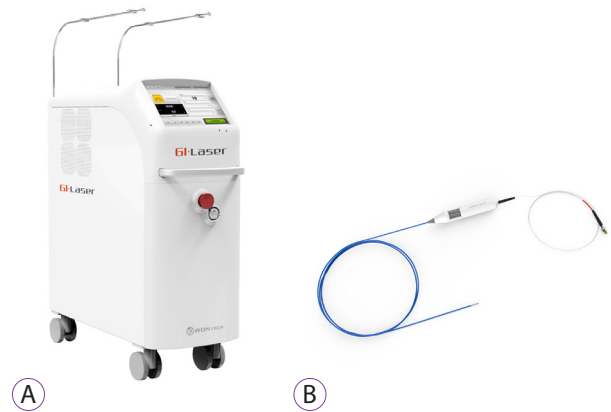


Fig. 1. Laser devices. (A) 1,940-nm wavelength diode laser system. (B) Fiber optic laser catheter.

and deliver the laser fiber to the surgical location through the working channel of the endoscope. The length of the fiber tip of this device can be adjusted by the operator according to the type of procedure, and up to 18 W of output can be delivered to the target tissue (Fig. 1).

Colonic endoscopic submucosal dissection using the fiber-guided diode laser system in the *ex vivo* porcine model

This experiment was exempt from institutional review board approval because no human research participants or live animals were involved. Fresh *ex vivo* porcine colorectal specimens, which included the rectum, sigmoid, and descending colon, were obtained from a commercial distributor. The length of the colon specimens was 40 cm, and their diameter ranged from 6 to 8 cm. The specimens were used in the EAS-IE-R simulator platform (EndoSim, LLC, Berlin, MA, USA).

Ten standardized artificial lesions measuring 3×3 cm were created using an endoscopic ruler approximately 15 cm from the anal verge in the porcine colorectal specimens. Thereafter, colonic ESD was performed using the FDLS by an experienced endoscopist who had performed more than 500 cases of human gastric or colorectal ESD (Supplementary Video 1).

Gastric and rectal endoscopic submucosal dissection using the fiber-guided diode laser system in the *in vivo* porcine model

Two mini pigs (*Sus scrofa*; Medikinetics Co., Ltd., Pyeongtaek, Korea; mean body weight, 30 kg; mean age, 14 months) were prepared for the *in vivo* experiment. All animal experiments were performed at the NCEED large animal endoscopy center after the experimental protocol was approved by the Institutional Animal Care Committee of KNOTUS Co.,

Ltd., a contract research organization for preclinical testing, before commencement of the study (KNOTUS IACUC 19-KE-359). Before testing, each animal was anesthetized with an intramuscular injection of tiletamine/zolazepam and xylazine. Eight ESD procedures (four gastric and four rectal ESD procedures) were performed by eight endoscopists who had performed more than 300 cases of human gastric or colorectal ESD (Supplementary Video 2).

Endoscopic submucosal dissection procedures

The procedures were performed using the FDLS and the following steps instead of conventional endoscopic knives: artificial lesions 3 cm in diameter were created at sites in the stomach or rectum easily reached by the endoscope by marking using the FDLS. A 0.9% normal saline solution combined with 0.05% indigo carmine was injected to the submucosal space at the target lesion. In the gastric ESD cases, a circumferential incision was created a few millimeters outside the margins of the target lesion. Thereafter, submucosal dissection was performed until the lesion was completely removed. In the colorectal ESD cases, a mucosal injection to the anal side of the tumor and a partial mucosal incision were performed. Submucosal dissection was then performed to prepare a mucosal flap. After the creation of the flap, half to two-thirds of the dissection was performed, followed by a full circumferential mucosal incision. Thereafter, submucosal dissection was performed until the lesion was completely removed.

Main outcome measurements

After we retrieved the excised lesion, the specimen and resection bed were inspected to determine the *en bloc* resection status. The specimens were spread and pinned on flat cork plates immediately after resection. The largest and perpendicular diameters of the specimen and resection area were measured. The procedure time, injection volume, and injection frequency were recorded, and the incidence of adverse events (bleeding and perforation) was monitored by an independent observer. The procedure speed, defined as the area of mucosa dissected per unit time (cm²/min), was calculated. All pigs were sacrificed after the procedures using excess isoflurane, and the stomach and colon of each pig were harvested for inspection and histopathological examination. These procedures were performed by a pathologist who was blinded to the study groups.

Perforations were assessed by searching for visible holes in endoscopic view or the harvested organs, using the underwater test, and by evaluating histopathologically. The degree of injury to the muscularis propria (MP) layer was histologically classified into four categories: grade 1 (mild), involving <50% of the inner circular layer; grade 2, involving >50% of the in-

ner circular layer but not reaching the outer longitudinal layer; grade 3, involving <50% of the outer longitudinal layer; and grade 4, involving >50% of the outer longitudinal layer and considered a histopathological perforation. The degree of injury to the MP layer per remaining tissue bed was recorded.¹⁴

Statistical analysis

Data were presented as means ± standard deviations and were evaluated for differences using the SPSS software, version 18.0 (SPSS Inc., Chicago, IL, USA). The independent samples *t*-test was used to compare the results between the gastric and colonic ESD groups. Categorical variables were compared using Fisher’s exact test. A *p*-value of <0.05 was considered significant.

RESULTS

Ex vivo animal experiments

All 10 colonic ESD procedures were completed using the FDLS without an endoscopic knife. The mean size of the resected specimens was 4.1 × 4.0 cm. The mean total procedure time and speed were 21.2 ± 7.3 (range, 13.5–34.3) min and 0.87 ± 0.31 (range, 0.38–1.31) cm²/min, respectively. The mean injection volume and number of MP layer injuries were 33.0 ± 11.8 mL and 0.8 ± 1.5, respectively. The *en bloc* and complete resection rates were both 100% (10/10), and perforation occurred in the first case only (Table 1). The mean injection volume was significantly larger (*p* = 0.046) in the first five cases than in the last five cases. The mean procedure time (25.2 ± 8.1 min vs. 40.8 ± 12.2 min) and number of MP layer injuries (0.2 ± 0.4 vs. 1.4 ± 1.9) tended to be shorter and lesser, respectively, in the last five cases than in the first five cases (*p* > 0.05) (Figs. 2 and 3).

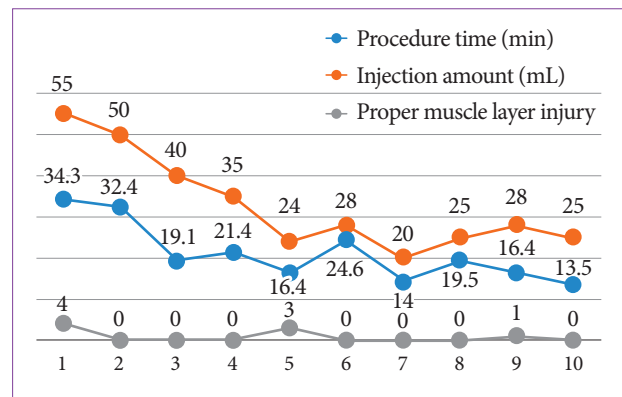


Fig. 2. Results of colonic endoscopic submucosal dissection in the *ex vivo* model.

Table 1. Results of Colonic Endoscopic Submucosal Dissection Using a Fiber-Guided Diode Laser System in the *ex vivo* Model

Cases	<i>En bloc</i>	R0 resection	Procedure time (min)	Procedure speed (cm ² /min)	Injection volume (mL)	Number of MP injuries	Perforation	Area of specimen (cm ²)
1	Yes	Yes	34.3	0.41	55	4	Yes	14
2	Yes	Yes	32.4	0.38	50	0	No	12.3
3	Yes	Yes	19.1	1.06	40	0	No	20.3
4	Yes	Yes	21.4	0.65	35	0	No	14
5	Yes	Yes	16.4	0.98	24	3	No	16
6	Yes	Yes	24.6	0.73	28	0	No	18
7	Yes	Yes	14.0	1.14	20	0	No	16
8	Yes	Yes	19.5	1.04	25	0	No	20.3
9	Yes	Yes	16.4	0.98	28	1	No	16
10	Yes	Yes	13.5	1.31	25	0	No	17.6
Total	100%	100%	21.2±7.3	0.87±0.31	33.0±11.8	0.8±1.5	10%	16.4±2.6

MP, muscularis propria.

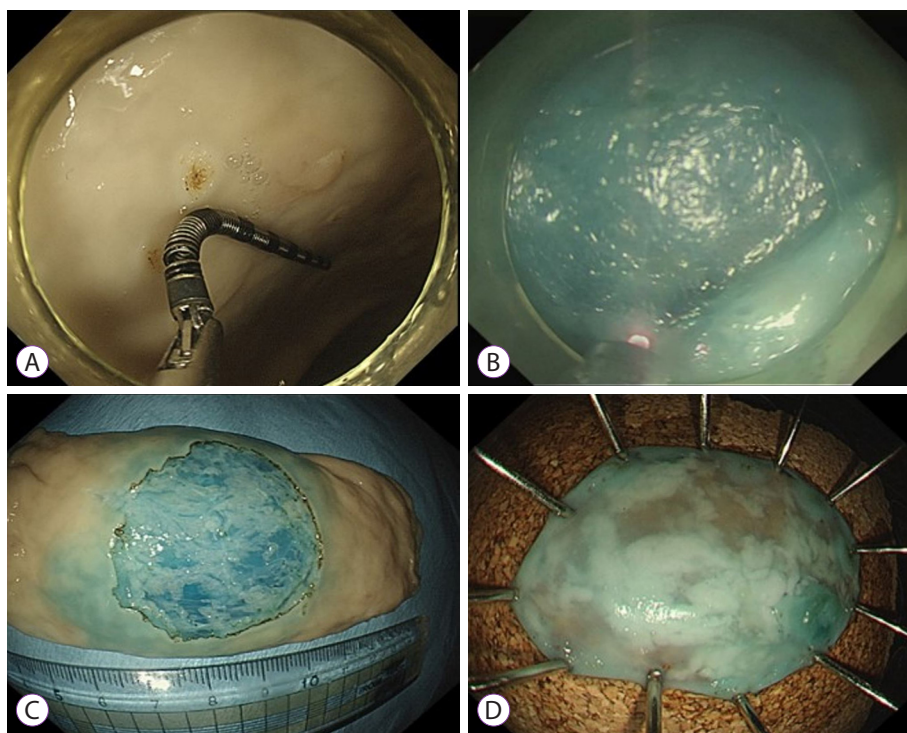


Fig. 3. Endoscopic submucosal dissection using the fiber-guided diode laser system in the *ex vivo* model. (A) Creation of artificial lesion 3 cm in diameter using an endoscopic ruler. (B) Submucosal dissection using the fiber optic laser catheter. (C) Remaining porcine colon tissue bed after endoscopic resection. (D) Specimen spread and pinned on cork plates immediately after resection.

In vivo animal experiments

Eight ESD procedures (four gastric ESD and four rectal ESD procedures) were completed via *en bloc* resection using the FDLS without an endoscopic knife. The mean marginal cutting time and submucosal dissection time were 4.5 ± 5.1 min and 21.0 ± 4.4 min, respectively, in the gastric ESD group. The

mean total procedure time, procedure speed, and specimen area were 27.7 ± 7.3 min, 0.42 ± 0.10 cm²/min, and 11.1 ± 1.3 cm² in the gastric ESD group and 28.5 ± 5.8 min, 0.42 ± 0.09 cm²/min, and 11.6 ± 1.2 cm² in the rectal ESD group (Tables 2 and 3).

Table 2. Results of Gastric and Rectal Endoscopic Submucosal Dissection Using a Fiber-Guided Diode Laser System in the *in vivo* Model

Case	Location	<i>En bloc</i>	Marginal cutting time (min)	Dissection time (min)	Total Procedure time (min)	Procedure speed (cm ² /min)	Injection volume (mL)	Injection frequency	Area of specimen (cm ²)	Grade of MP injury	Perforation	Bleeding
1	Stomach	Yes	1.2	25.2	26.4	0.41	26	10	10.9	1	No	No
2	Stomach	Yes	1	17.5	18.5	0.55	20	13	10.2	0	No	No
3	Stomach	Yes	4	17	30	0.34	22	16	10.2	0	No	No
4	Stomach	Yes	11.8	24.3	36.1	0.35	20	12	12.9	1	No	No
5	Rectum	Yes			32.1	0.41	20	18	13.3	2	No	No
6	Rectum	Yes			29.9	0.39	6	3	11.5	0	No	No
7	Rectum	Yes			32	0.34	30	28	10.8	4	Yes	No
8	Rectum	Yes			20	0.54	10	9	10.9	1	No	No

MP, muscularis propria.

Table 3. Results of Gastric and Rectal Endoscopic Submucosal Dissection in the *in vivo* Model

	Gastric ESD group (n=4)	Rectal ESD group (n=4)
Marginal cutting time, min	4.5±5.1	
Submucosal dissection time, min	21±4.4	
Total procedure time, min	27.7±7.3	28.5±5.8
Procedure speed, cm ² /min	0.42±0.10	0.42±0.09
Area of specimen (mean±SD), cm ²	11.1±1.3	11.6±1.2
<i>En bloc</i> , %	100 (4/4)	100 (4/4)
Intraoperative bleeding points (n)	0	0
Injection frequency (mean±SD)	12.8±2.5	14.5±10.9
Injection volume (mean±SD), mL	22±2.8	16.5±10.8
Grade of MP injury		
1	2	1
2	0	1
3	0	0
4	0	1
Perforation	0	25% (1/4)

ESD, endoscopic submucosal dissection; MP; muscularis propria; SD, standard deviation.

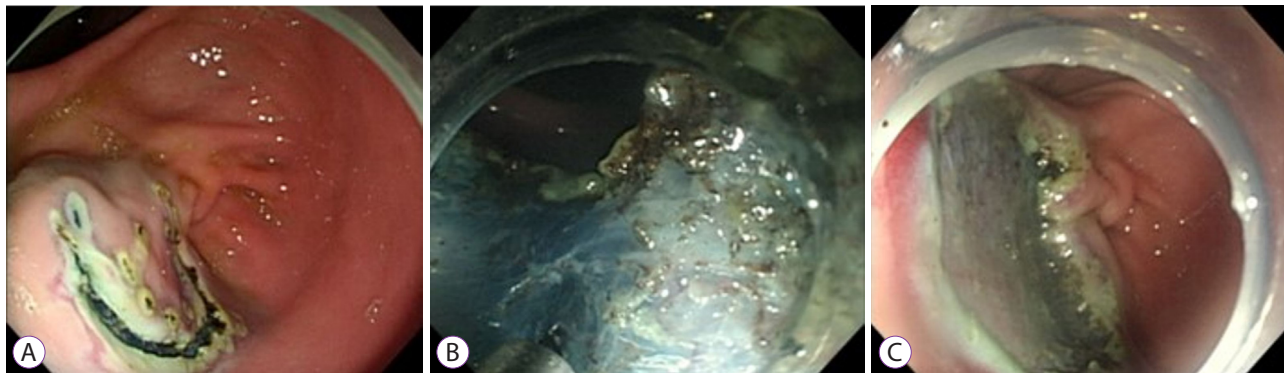


Fig. 4. Endoscopic submucosal dissection using the fiber-guided diode laser system in the *in vivo* model. (A) Circumferential incision performed outside the margins of the target lesion. (B) Submucosal dissection using the fiber optic laser catheter. (C) Completely resected target lesion.

In the gastric ESD group, no MP layer injury occurred in 50% (2/4) of the cases, and grade 1 MP layer injury occurred in 50% (2/4) of the cases. In the rectal ESD group, MP layer injury did not occur in 25% (1/4) of the cases; however, grade 1, 2, and 4 MP layer injuries occurred in 25% (1/4) of the cases each. One case of perforation occurred only in the rectal ESD group. No intraprocedural bleeding occurred in either group (Fig. 4).

DISCUSSION

ESD is one of the most technically difficult endoscopic procedures. According to the recently updated guidelines of the Japan Gastroenterological Endoscopy Society, understanding anatomical features and familiarity with the basic techniques of polypectomy, endoscopic mucosal resection, endoscopic hemostasis, and clip suture are the minimum requirements before starting ESD.¹⁵ Novices must practice using *ex vivo* or *in vivo* animal models or in the clinic under a supervisor to perform ESD more safely and efficiently. Several animal and clinical trials reported that at least 30 cases of gastric ESD should be performed to achieve an acceptable technical level.^{16,17} However, one study reported that a high technical level cannot be reached until more than 240 cases of colorectal ESD have been performed.¹⁸

Several factors increase the difficulty of ESD, including intraprocedural bleeding, paradoxical scope control, non-lifting lesions, perpendicular submucosal approach, ineffective gravity countertraction, and submucosal fibrosis.¹⁹ The incidence of intraprocedural bleeding during ESD is >20%,²⁰ and proper management of bleeding plays a critical role in successful ESD because it interferes with precise cutting under clear vision.

Several studies have evaluated the clinical effectiveness and applicability of various types of laser devices capable of precise tissue cutting without intraprocedural bleeding during ESD. Cho et al.¹³ reported preliminary results of gastric ESD using thulium laser. Ten patients underwent ESD using thulium laser without an endoscopic knife, and the curative resection rate was 90% (9/10). No active intraprocedural bleeding, delayed bleeding, or perforation was detected. Preliminary results of gastric ESD were also reported with the use of CO₂ laser in *ex vivo* and *in vivo* porcine models. *En bloc* resection was achieved in all eight gastric ESD (five *ex vivo* and three *in vivo*) cases, and there was no perforation, MP layer injury, or uncontrollable hemorrhage observed during the procedures.¹¹

The diode laser system has been used for a variety of surgeries, such as prostate disease surgery,²¹ endovenous ablation treatment,²² tonsillectomy,²³ and laser-assisted laparoscopic partial nephrectomy,²⁴ and has proven to be effective for tissue excision and hemostasis. Tang et al.¹⁴ demonstrated the feasibility of ESD using diode laser in an animal model. Eight cases of esophageal ESD were performed using the synchronous 980/1,470-nm dual-diode laser system, achieving a 100% *en bloc* resection rate without perforation. The procedure speed of ESD was significantly faster using the dual-diode laser system than using a conventional endoscopic knife (0.27 cm²/min vs. 0.21 cm²/min, *p* = 0.001). In addition, fewer intraprocedural bleeding points were detected, and the use of hemostatic forceps was significantly less frequent in the diode laser ESD group than in the conventional ESD group. However, studies on diode laser-assisted ESD are very rare. The 1,940-nm diode laser system, as demonstrated in tonsillectomy, is capable of precise excision of the mucosa and submucosa without intraoperative bleeding; however, no studies related to endoscopic procedures have been published.²³ To apply these advantages

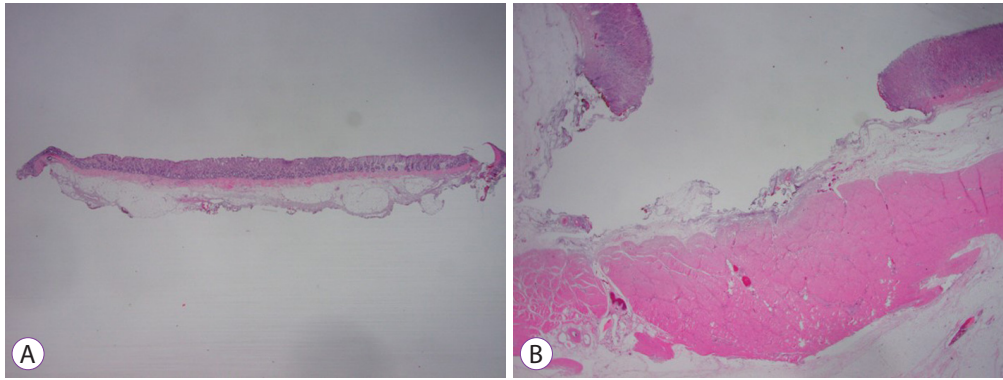


Fig. 5. Histopathological examination in the *in vivo* study. (A) Resected colon tissue specimen (hematoxylin and eosin staining, $\times 12.5$ magnification), (B) remainder of the stomach after endoscopic resection (hematoxylin and eosin staining, $\times 12.5$ magnification).

of diode laser to ESD, we conducted gastric or colorectal ESD using a specially designed fiber optic laser catheter that enables tissue cutting through the endoscope working channel, similar to an endoscopic knife.

In our *ex vivo* study, *en bloc* and complete resections were achieved in all 10 cases of colonic ESD using the FDLS. Perforation occurred in the first case of colonic ESD because it was initially difficult to maintain the proper cutting direction using the FDLS. Afterwards, only minor MP layer injuries occurred, without any perforations in the remaining cases. As the operator adapted relatively quickly, the number of MP layer injuries (0.2 ± 0.4 vs. 1.4 ± 1.9) and procedure time (25.2 ± 8.1 min vs. 40.8 ± 12.2 min) were lower and shorter, respectively, and the procedure speed (1.04 ± 0.21 cm²/min vs. 0.70 ± 0.31 cm²/min) was faster in the last five cases than in the first five cases.

In our *in vivo* study, the *en bloc* resection and perforation rates were 100% and 12.5%, respectively, although most endoscopists had no experience with ESD using the FDLS. No intraprocedural bleeding occurred; however, MP layer injuries were more frequent during rectal ESD (25% grade 1, 25% grade 2, and 25% grade 4 injuries) than during gastric ESD (50% grade 1 injury). These results probably occurred because the colon wall is thinner than the stomach wall, and the colon lumen is narrower than the stomach lumen, making endoscopic manipulation difficult.

Based on the results of our study, FDLS-guided ESD has some advantages, such as a low risk of bleeding and safe cutting in the area of submucosal injection. Intraprocedural bleeding did not occur in this study. If it had occurred, it is expected that hemostasis would have been achieved with the FDLS using the non-contact technique without changing to other hemostasis devices. The FDLS enabled cutting the mucosa or submucosa very safely after the submucosal injection

because the output wavelength of a 1,940-nm laser coincides with the absorption peak of water. In addition, the high absorption coefficient of a 1,940-nm laser allows a shallow penetration depth, enabling precise ablation with less thermal damage.^{23,25} When the resected specimens were evaluated histologically, the cutting surface was observed accurately enough to judge whether the lesion was completely excised (Fig. 5).

However, there were some disadvantages when using this device. The FDLS-guided ESD technique differs from the conventional ESD technique. Conventional needle-type ESD knives enable tissue resection of any tissue in contact with the metal tip; however, the apex of the tip can be used to resect the tissue when using the fiber-optic laser catheter. This limitation made it technically difficult to secure an appropriate resection plane when the lesion was in a perpendicular position. Thus, it was important to maintain a safe resection plane parallel to the MP layer during submucosal dissection.

Some limitations of this study should be discussed. The sample size was small, and we did not reflect the condition of the patients' breathing or heartbeat. We selected the optimal location for the ESD procedure and did not compare it with the use of conventional endoscopic knives. Despite these limitations, this is the first study to investigate the usefulness of the 1,940-nm FDLS for gastric and colorectal ESD. Large animal trials or human clinical trials are needed to confirm the advantages of this device over conventional endoscopic knives.

In conclusion, the 1,940-nm FDLS was capable of yielding a high *en bloc* resection rate without intraprocedural bleeding during gastric and colorectal ESD in an animal model.

Conflicts of Interest

Wontech Co., Ltd. (Daejeon, Korea) provided the FDLS and fiber optic laser catheter. The authors have no potential conflicts of interest.

Funding

None.

Author Contributions

Conceptualization: Yunho Jung, Gwang Ho Baik, Weon Jin Ko, Joo Young Cho

Data curation: YJ, GHB, WJK, Bong Min Ko, Seong Hwan Kim, Jin Seok Jang, Jae-Young Jang, Wan-Sik Lee, Young Kwan Cho, Sun Gyo Lim, Hee Seok Moon, In Kyung Yoo, JYC

Formal analysis: YJ

Investigation: BMK, SHK, JSJ, JYJ, WSL, YKC, SGL, HSM, IKY

Methodology: JYC

Project administration: GHB, JYC

Resources: WJK, JYC

Writing-original draft: YJ, JYC

Writing-review&editing: GHB

ORCID

Yunho Jung:	https://orcid.org/0000-0002-7760-0050
Gwang Ho Baik:	https://orcid.org/0000-0003-1419-7484
Weon Jin Ko:	https://orcid.org/0000-0002-3796-8539
Bong Min Ko:	https://orcid.org/0000-0002-0635-4454
Seong Hwan Kim:	https://orcid.org/0000-0002-1276-3334
Jin Seok Jang:	https://orcid.org/0000-0001-8067-4598
Jae-Young Jang:	https://orcid.org/0000-0002-7930-1468
Wan-Sik Lee:	https://orcid.org/0000-0002-8021-6228
Young Kwan Cho:	https://orcid.org/0000-0001-8042-3828
Sun Gyo Lim:	https://orcid.org/0000-0003-2045-5099
Hee Seok Moon:	https://orcid.org/0000-0002-8806-2163
In Kyung Yoo:	https://orcid.org/0000-0003-0909-339X
Joo Young Cho:	https://orcid.org/0000-0002-7182-5806

Supplementary Materials

Video 1. *Ex vivo* colon endoscopic submucosal dissection using fiber-guided diode laser system (<https://doi.org/10.5946/ce.2020.229.v001>).

Video 2. *In vivo* gastric endoscopic submucosal dissection using fiber-guided diode laser system (<https://doi.org/10.5946/ce.2020.229.v002>).

REFERENCES

1. Bhatt A, Abe S, Kumaravel A, Vargo J, Saito Y. Indications and techniques for endoscopic submucosal dissection. *Am J Gastroenterol* 2015;110:784-791.
2. Belderbos TD, Leenders M, Moons LM, Siersema PD. Local recurrence after endoscopic mucosal resection of nonpedunculated colorectal lesions: systematic review and meta-analysis. *Endoscopy* 2014;46:388-402.
3. Ko BM. History and development of accessories for endoscopic submucosal dissection. *Clin Endosc* 2017;50:219-223.
4. Horikawa Y, Fushimi S, Sato S. Hemorrhage control during gastric endoscopic submucosal dissection: techniques using uncovered knives. *JGH Open* 2020;4:4-10.
5. Goto O, Fujishiro M, Oda I, et al. A multicenter survey of the management after gastric endoscopic submucosal dissection related to postoperative bleeding. *Dig Dis Sci* 2012;57:435-439.
6. Park YM, Cho E, Kang HY, Kim JM. The effectiveness and safety of endoscopic submucosal dissection compared with endoscopic mucosal resection for early gastric cancer: a systematic review and metaanalysis. *Surg Endosc* 2011;25:2666-2677.

7. Jang JS, Choi SR, Graham DY, et al. Risk factors for immediate and delayed bleeding associated with endoscopic submucosal dissection of gastric neoplastic lesions. *Scand J Gastroenterol* 2009;44:1370-1376.
8. Reumkens A, Rondagh EJ, Bakker CM, Winkens B, Masclee AA, Sanduleanu S. Post-colonoscopy complications: a systematic review, time trends, and meta-analysis of population-based studies. *Am J Gastroenterol* 2016;111:1092-1101.
9. Kichler A, Jang S. Endoscopic hemostasis for non-variceal upper gastrointestinal bleeding: new frontiers. *Clin Endosc* 2019;52:401-406.
10. Noguchi T, Hazama H, Nishimura T, Morita Y, Awazu K. Enhancement of the safety and efficacy of colorectal endoscopic submucosal dissection using a CO₂ laser. *Lasers Med Sci* 2020;35:421-427.
11. Obata D, Morita Y, Kawaguchi R, et al. Endoscopic submucosal dissection using a carbon dioxide laser with submucosally injected laser absorber solution (porcine model). *Surg Endosc* 2013;27:4241-4249.
12. Saccomandi P, Quero G, Costamagna G, Diana M, Marescaux J. Effects of Nd:YAG laser for the controlled and localized treatment of early gastrointestinal tumors: preliminary in vivo study. *Annu Int Conf IEEE Eng Med Biol Soc* 2017;2017:4533-4536.
13. Cho JH, Cho JY, Kim MY, et al. Endoscopic submucosal dissection using a thulium laser: preliminary results of a new method for treatment of gastric epithelial neoplasia. *Endoscopy* 2013;45:725-728.
14. Tang J, Ye S, Ji X, Li J, Liu F. Comparison of synchronous dual wavelength diode laser versus conventional endo-knives for esophageal endoscopic submucosal dissection: an animal study. *Surg Endosc* 2018;32:5037-5043.
15. Tanaka S, Kashida H, Saito Y, et al. Japan gastroenterological endoscopy society guidelines for colorectal endoscopic submucosal dissection/endoscopic mucosal resection. *Dig Endosc* 2020;32:219-239.
16. Kato M, Gromski M, Jung Y, Chuttani R, Matthes K. The learning curve for endoscopic submucosal dissection in an established experimental setting. *Surg Endosc* 2013;27:154-161.
17. Yoshida M, Kakushima N, Mori K, et al. Learning curve and clinical outcome of gastric endoscopic submucosal dissection performed by trainee operators. *Surg Endosc* 2017;31:3614-3622.
18. Rönnow CF, Uedo N, Toth E, Thorlaciuc H. Endoscopic submucosal dissection of 301 large colorectal neoplasias: outcome and learning curve from a specialized center in Europe. *Endosc Int Open* 2018;6:E1340-E1348.
19. Iacopini F, Saito Y, Bella A, et al. Colorectal endoscopic submucosal dissection: predictors and neoplasm-related gradients of difficulty. *Endosc Int Open* 2017;5:E839-E846.
20. Oka S, Tanaka S, Kaneko I, et al. Advantage of endoscopic submucosal dissection compared with EMR for early gastric cancer. *Gastrointest Endosc* 2006;64:877-883.
21. Zhang J, Li J, Wang X, Shi C, Tu M, Shi G. Efficacy and safety of 1470-nm diode laser enucleation of the prostate in individuals with benign prostatic hyperplasia continuously administered oral anticoagulants or antiplatelet drugs. *Urology* 2020;138:129-133.
22. Aktas AR, Celik O, Ozkan U, et al. Comparing 1470- and 980-nm diode lasers for endovenous ablation treatments. *Lasers Med Sci* 2015;30:1583-1587.
23. Kang K, Kim H, Kim JH, et al. The feasibility of 1940-nm diode laser in tonsillectomy. *Medical Lasers* 2016;5:77-82.
24. Khoder WY, Sroka R, Siegert S, Stief CG, Becker AJ. Outcome of laser-assisted laparoscopic partial nephrectomy without ischaemia for peripheral renal tumours. *World J Urol* 2012;30:633-638.
25. Guney M, Tunc B, Gulsoy M. Investigating the ablation efficiency of a 1940-nm thulium fibre laser for intraoral surgery. *Int J Oral Maxillofac Surg* 2014;43:1015-1021.